

# Community baseline vertebrate biodiversity surveys in the Hornsby Council region using eDNA

## Friday, 26 July 2024

Project number:	ED_2406CR1
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Assay(s):	<i>Vertebrate (12S), Decapod</i>
Filter used:	1.2 µm EnviroDNA manual syringe disc filter

## Highlights

- At each of the 41 sites, 2 water samples were collected in Spring 2023 and Autumn 2024.
- Spring 2023 results:
  - o 67% (vertebrate assay) and 80% (decapod assay) of taxa were resolved at the species level.
  - o Across all 41 sites, 97 vertebrate taxa, including 1 threatened species (*Pteropus poliocephalus*) and 5 decapod taxa were detected.
  - o Taxon richness at the site level ranged from 3 to 29 (vertebrate assay) and from 0 to 2 (decapod assay).
  - o Longfin eel (*Anguilla reinhardtii*) (vertebrate assay) and Sydney spiny crayfish (*Euastacus australasiensis*) (decapod assay) was the most commonly detected taxon.
  - o >5000 reads were obtained for 96% (vertebrate assay) and 41% (decapod assay) of metabarcoding samples.
- Autumn 2024 results:
  - o 78% (vertebrate assay) and 80% (decapod assay) of taxa were resolved at the species level.
  - o Across all sites, 79 vertebrate taxa, including 3 threatened species; Powerful owl (*Ninox strenua*), Grey-headed flying-fox (*Pteropus poliocephalus*) and Pilotbird, (*Pycnoptilus floccosus*) and 5 decapod taxa were detected.
  - o Taxon richness at the site level ranged from 0 to 17 (vertebrate assay) and from 0 to 2 (decapod assay).
  - o Longfin eel (*Anguilla reinhardtii*) (vertebrate assay) and Sydney Spiny Crayfish (*Euastacus australasiensis*) (decapod assay) was the most commonly detected taxon.

## Background

Environmental DNA (eDNA) methods are being used routinely to monitor aquatic animals including fish, amphibians and mammals across waterways, estuaries and wetlands throughout Australian catchments. Here we use both a vertebrate and decapod eDNA metabarcoding assay to screen 82 eDNA samples taken from 41 sites throughout the Hornsby council region, New South Wales to provide a baseline biodiversity assessment of vertebrate and decapod species during Spring 2023 and Autumn 2024.

## Methods

### Sampling

During both Spring 2023 and Autumn 2024, 82 water samples were collected from 41 sites (Figure 1) by Hornsby council staff and citizen scientists. At each site, 2 replicate samples were collected by passing up to 2,000 mL of water (mean = 907 mL) in the Spring sampling round and 4,000 mL (mean = 1513 mL) in the Autumn sampling round through a 1.2 µm EnviroDNA manual syringe disc filter. Filtration was undertaken on-site to reduce DNA degradation during transport of water samples. Filters were stored out of sunlight and at ambient temperature before being transported to the laboratory for processing.

### Analysis

DNA was extracted from filters using a Qiagen PowerSoil Kit that minimises compounds that can inhibit PCR reactions in environmental samples. Library construction involved two rounds of PCR, whereby the first round employed gene-specific primers to amplify the target region and the second round incorporated sequencing adapters and unique barcodes for each sample-amplicon combination included in the library. Negative controls were included during library construction. Negative controls consisted of the extraction negative as well as PCR negatives, in which nuclease-free water was used in place of DNA during both rounds of PCR. Sequencing was carried out on an Illumina sequencing platform.

Following quality control filtering to remove primer sequences, truncated reads, and low-frequency reads, DNA sequences were clustered into Operational Taxonomic Units (OTUs) on the basis of sequence similarity. Taxonomic assignment was performed with VSEARCH software (Rognes et al. 2016), whereby each OTU cluster was assigned a species identity using a threshold of 95% by comparing against a reference sequence database. Where a species could not be assigned (i.e., reference database was deficient and/or taxa were poorly-characterised), taxonomic assignments were manually vetted by first obtaining a list of possible species through BLASTN searches against the public repository Genbank ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)), followed by elimination of species on the basis of their geographic distributions, using information from the Atlas of Living Australia and other relevant data sources. In cases where an OTU could not be adequately resolved to a single species (e.g., due to shared haplotypes), either a list of multiple species is included, or the OTU is assigned to the lowest taxonomic rank without further classification.

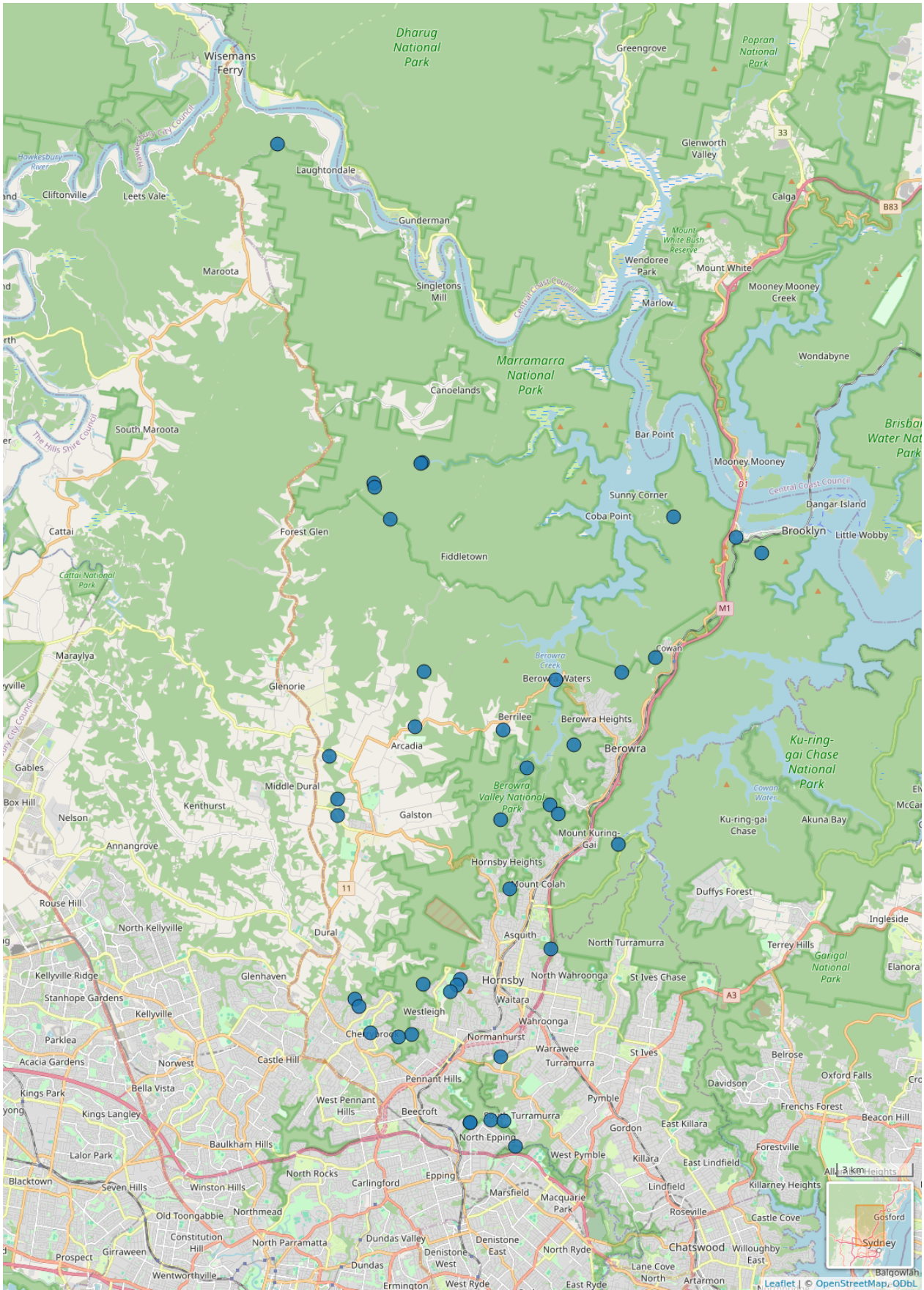


Figure 1. Locations of the 41 sites sampled in both Spring 2023 and Autumn 2024 in the Hornsby Council region, NSW.

## Results

A total of 82 samples were analysed from 41 sites across the Hornsby Council region, NSW using a 1.2 µm EnviroDNA manual syringe disc filter. Raw data on per-sample detections can be found in accompanying spreadsheet (ED\_2406CR1\_Hornsby\_VertDeca\_Data). The spreadsheet provides the taxa detected in each sample, as well as the number of sequence reads for each taxon. Reads should not be directly interpreted as taxa abundance. While some studies have shown a positive correlation between read numbers and abundance, reads can also be influenced by a number of other variables. Reads may be used to help assign a level of confidence in species detection along with the number of replicates in which the species was detected.

### Spring 2023 sampling results

#### Vertebrate Metabarcoding

Overall, 97 vertebrate taxa were detected, including 13 introduced species and 1 species listed at the Federal and/or State level; Grey-headed flying-fox (*Pteropus poliocephalus*) (listing data from <https://www.environment.gov.au/sprat-public/action/report>). Six frog, 25 fish, 6 reptile, 36 bird, and 24 mammal taxa were detected. The number of vertebrate taxa at each site (across all replicate samples) ranged from 3 to 29. The number of native taxa per site varied from 1 to 16.

Most taxa were resolved at the species level (vertebrate assay = 67%, decapod assay = 80% of all taxa). The fact that some taxa could not be resolved at the species level is likely due to inadequate genetic sequence data available in the reference library for the region. Further reference sequences for species that are not currently captured in the reference database are needed to fully evaluate the potential for the 12S region to resolve these taxa to a species or genus level. Unresolved taxa can also arise due to limitations with the target region (e.g., 12S, 16S) and metabarcoding assays in general, whereby only a very small subset of the entire genome is interrogated for the purpose of species identification. Consequently, there is not always enough genetic variation in that short marker sequence to definitively assign it to a species.

A summary of the frequency of occurrence of each vertebrate species across all samples and sites is provided in Table 1.

Table 1. Number of detections and number of occupied sites for each vertebrate taxon during the Spring 2023 sampling season.

Group	Taxa	Common name	N detections	N sites
	<i>Acanthagenys rufogularis</i>	Spiny-cheeked honeyeater	1	1
	<i>Acanthiza pusilla</i>	Brown thornbill	2	2
	<i>Alectura lathamii</i>	Australian brush-turkey	3	3
Birds	<i>Alisterus scapularis</i>	Australian king-parrot	3	3
	<i>Anatidae</i>	Family of waterbirds that includes ducks, geese and swans	6	5
	<i>Cacatua</i>	Genus of cockatoo	1	1



Group	Taxa	Common name	N detections	N sites
	<i>Cacatua galerita</i>	Sulphur-crested cockatoo	13	10
	<i>Charadriiformes</i>	Order of shorebirds	1	1
	<i>Chenonetta jubata</i>	Australian wood duck	1	1
	<i>Columba livia</i>	Domestic pigeon	3	3
	<i>Cormobates leucophaea</i>	White-throated treecreeper	2	2
	<i>Corvus</i>	Genus of crows and ravens	1	1
	<i>Dacelo novaeguineae</i>	Laughing kookaburra	6	4
	<i>Eolophus roseicapilla</i>	Galah	2	2
	<i>Gallinula tenebrosa</i>	Dusky moorhen	6	4
	<i>Malurus lamberti</i>	Variiegated fairy-wren	1	1
	<i>Manorina</i>	Genus of australian honeyeaters and miners	9	6
	<i>Meliphaga lewinii</i>	Lewin's honeyeater	1	1
	<i>Meliphagidae</i>	Family of honeyeaters	4	3
	<i>Menura novaehollandiae</i>	Superb lyrebird	1	1
	<i>Microcarbo melanoleucos</i>	Little pied cormorant	5	4
	<i>Ocyphaps lophotes</i>	Crested pigeon	1	1
	<i>Pachycephala</i>	Genus of whistlers	1	1
	<i>Passeriformes</i>	Order of perching birds	8	7
	<i>Phalacrocorax sulcirostris</i>	Little black cormorant	4	3
	<i>Platycercus eximius</i>	Eastern rosella	9	8
	<i>Podargus strigoides</i>	Tawny frogmouth	2	2
	<i>Poodytes</i>	Genus of grassbirds	2	2
	<i>Ptilonorhynchus violaceus</i>	Satin bowerbird	1	1
	<i>Sericornis frontalis</i>	White-browed scrubwren	1	1
	<i>Sturnidae</i>	Family of starlings	1	1
	<i>Sturnus vulgaris</i>	Common starling	1	1
	<i>Trichoglossus</i>	Genus of lorikeet	6	6
	<i>Turdus</i>	Genus of thrush	4	3
	<i>Turdus philomelos</i>	Song thrush	2	2
	<i>Zanda</i>	Genus of black cockatoo	2	1
Fishes & eels	<i>Actinopteri</i>	Class of fish, unassigned	14	9
	<i>Anguilla</i>	Genus of freshwater eels	34	21
	<i>Anguilla reinhardtii</i>	Longfin eel	60	35
	<i>Carassius auratus</i>	Goldfish	3	3
	<i>Clupeidae</i>	Family of herrings and sprats	1	1
	<i>Cyprinus carpio</i>	European carp	7	4
	<i>Galaxias</i>	Genus of galaxiids	17	11

Group	Taxa	Common name	N detections	N sites
	<i>Gambusia</i>	Genus of mosquitofish	10	6
	<i>Gobiomorphus australis</i>	Striped gudgeon	24	13
	<i>Gobiomorphus coxii</i>	Cox's gudgeon	48	30
	<i>Gracilimugil argenteus</i>	Flat-tail mullet	2	1
	<i>Hypseleotris</i>	Genus of carp gudgeons	5	3
	<i>Melanotaenia</i>	Genus of rainbowfish	1	1
	<i>Mugil cephalus</i>	Sea mullet	2	1
	<i>Mugilidae</i>	Family of mullet	1	1
	<i>Mugilogobius platynotus</i>	Flatback mangrovegoby	1	1
	<i>Notesthes robusta</i>	Bullrout	2	1
	<i>Percalates novemaculeata</i>	Australian bass	14	8
	<i>Philypnodon grandiceps</i>	Flatheaded gudgeon	7	4
	<i>Philypnodon macrostomus</i>	Dwarf flathead gudgeon	2	1
	<i>Retropinna semoni</i>	Australian smelt	10	6
	<i>Salmo</i>	Genus of salmon and trout	1	1
	<i>Scomber</i>	Genus of mackerels	1	1
	<i>Tandanus tandanus</i>	Eel-tailed catfish, freshwater catfish	2	2
	<i>Tetractenos</i>	Genus of toadfish	2	1
Frogs	<i>Crinia signifera</i>	Common froglet	34	22
	<i>Limnodynastes peronii</i>	Striped marsh frog	23	14
	<i>Litoria fallax</i>	Eastern dwarf tree frog	3	2
	<i>Litoria gracilentata</i>	Dainty green tree frog	1	1
	<i>Litoria peronii</i>	Peron's tree frog	12	10
	<i>Litoria phyllochroa</i>	Leaf green tree frog	40	25
Mammals	<i>Bos taurus</i>	Cattle	8	7
	<i>Canis lupus</i>	Dog/dingo	18	14
	<i>Diprotodontia</i>	Order of marsupials that includes kangaroos, wallabies, possums	15	12
	<i>Eptesicus vulturinus</i> or <i>Vespadelus vulturinus</i>	Little forest bat	1	1
	<i>Hydromys chrysogaster</i>	Rakali	3	2
	<i>Macropodidae</i>	Family of marsupials that includes kangaroos, wallabies	3	2
	<i>Macropus giganteus</i>	Eastern grey kangaroo	1	1
	<i>Mormopterus planiceps</i>	Southeastern free-tailed bat	1	1
	<i>Mus musculus</i>	House mouse	4	4
	<i>Myotis adversus</i>	Large foot bat	1	1
<i>Notamacropus</i>	Genus of wallaby	1	1	

Group	Taxa	Common name	N detections	N sites
	<i>Oryctolagus cuniculus</i>	European rabbit	2	2
	<i>Ovis aries</i>	Sheep	4	3
	<i>Perameles</i>	Genus of bandicoots	1	1
	<i>Petaurus breviceps</i>	Sugar glider	2	2
	<i>Pseudocheiridae</i>	Family of ring-tailed possums	5	4
	<i>Pseudocheirus peregrinus</i>	Eastern ring-tailed possum	1	1
	<i>Pteropus poliocephalus</i>	Grey-headed flying-fox	9	7
	<i>Rattus</i>	Genus of rodents	5	5
	<i>Rattus fuscipes</i>	Bush rat	5	4
	<i>Rattus norvegicus</i>	Brown rat	3	2
	<i>Rattus rattus</i>	Black rat	9	8
	<i>Sus scrofa</i>	Pig	11	9
	<i>Trichosurus vulpecula</i>	Common brush-tailed possum	6	6
	<i>Chelodina</i>	Genus of snake necked turtles	1	1
	<i>Concinnia tenuis</i>	Barred-sided skink	3	2
	<i>Eulamprus quoyii</i>	Eastern water skink	8	6
Reptiles	<i>Myuchelys latisternum</i>	Saw-shelled turtle	1	1
	<i>Saproscincus mustelinus</i>	Weasel skink	4	3
	<i>Scincidae</i>	Genus of skinks	2	1

Figure 2, below, shows similar data to those presented in the table above. Rather than focusing on the number of detections, however, this figure shows the percentage of reads assigned to each taxon.



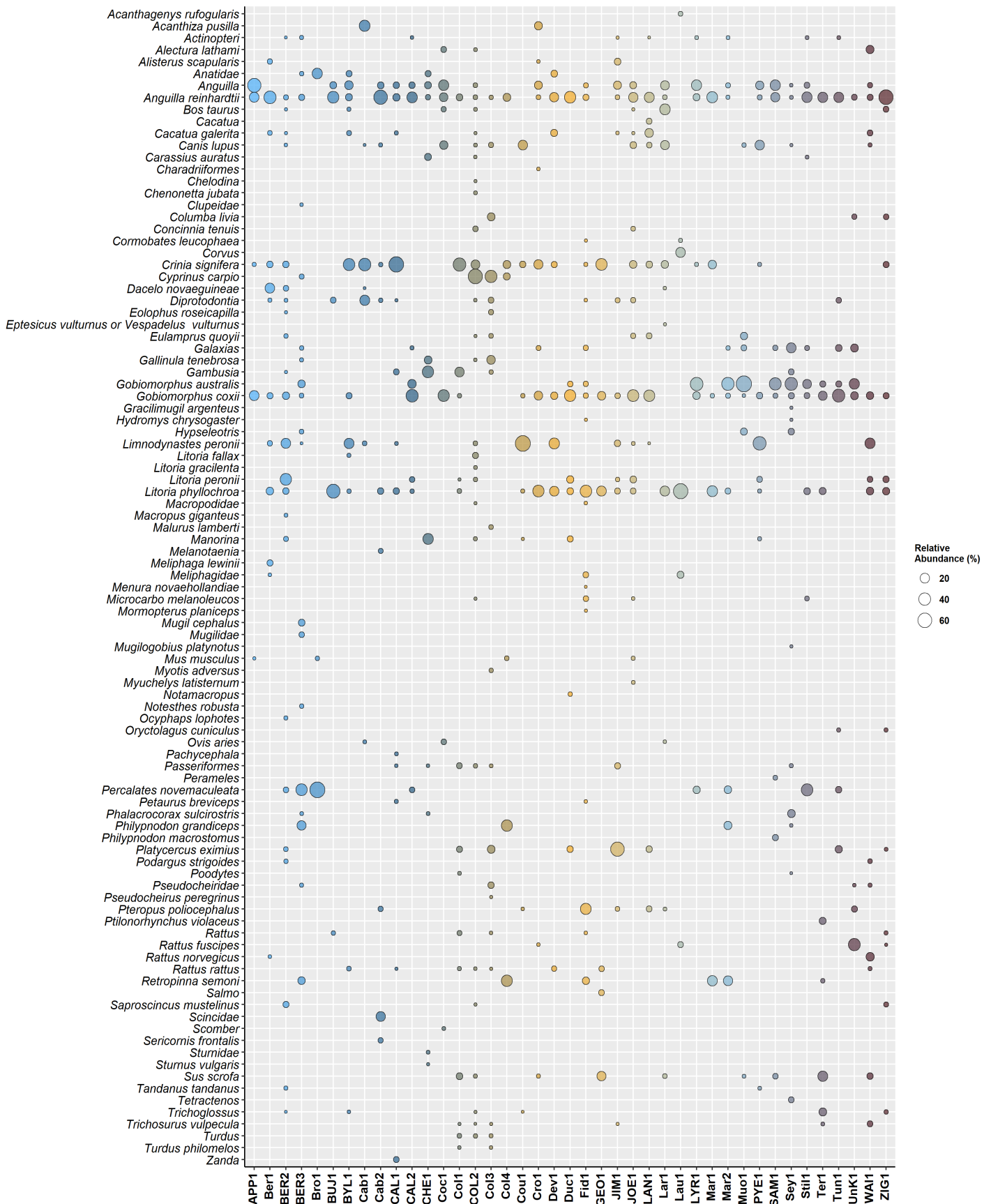


Figure 2. Percentage of reads assigned to each vertebrate taxon at each site in Spring 2023.

### Decapod Metabarcoding

Overall, five decapod taxa were detected. The number of decapod taxa at each site (across all replicate samples) ranged from 0 to 2.

The lower detection levels present in the decapod assay in comparison to the vertebrate assay may be due to factors such as differences in eDNA shedding and decay rates as well as high environmental flows and diluted DNA concentration present in the waterways.

A summary of the frequency of occurrence of each decapod species across the 41 sites is provided in Table 2.

Table 2. Number of detections and number of occupied sites for each decapod taxon during the Spring 2023 sampling season.

Family	Species	Common name	N detections	N sites
Atyidae	<i>Australatya striolata</i>	Riffle shrimp	2	2
	<i>Paratya australiensis</i>	Australian glass shrimp	7	6
Parastacidae	<i>Euastacus</i>	Genus of freshwater crayfish	3	2
	<i>Euastacus australasiensis</i>	Sydney spiny crayfish	4	3
	<i>Euastacus spinifer</i>	Giant spiny crayfish	7	6

The figure below shows similar data to those presented in the table above. Rather than focusing on the number of detections, however, this figure shows the percentage of reads assigned to each taxon in a sample.

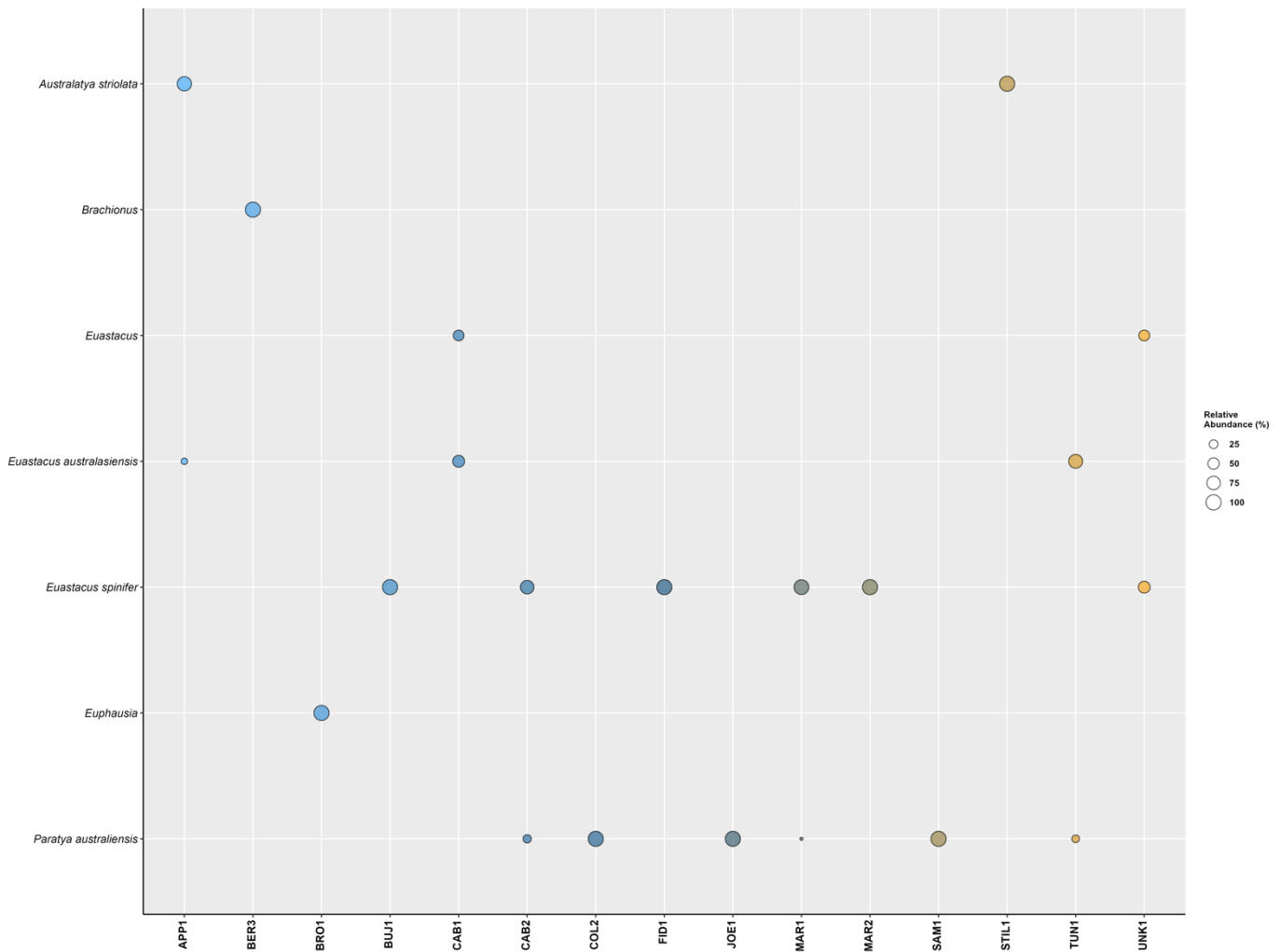


Figure 3. Percentage of reads assigned to each decapod taxon at each site in Spring 2023.

## Autumn 2024 sampling results

Most taxa were resolved at the species level (vert assay = 78%; decapod assay = 80% of all taxa).

### Vertebrate Metabarcoding

Overall, 79 vertebrate taxa were detected, including 12 vertebrate species that are not native to Australia and 3 vertebrate species Powerful owl (*Ninox strenua*), Grey-headed flying-fox (*Pteropus poliocephalus*) and Pilotbird, (*Pycnoptilus floccosus*) listed at the Federal and/or State level (listing data from <https://www.environment.gov.au/sprat-public/action/report>). Three frog, 19 fish, 4 reptile, 34 bird, and 19 mammal taxa were detected. The number of vertebrate taxa at each site (across all replicate samples) ranged from 1 to 17. The number of native taxa per site varied from 0 to 15.

A summary of the frequency of occurrence of each vert species across all samples and sites is provided in Table 1.

Table 3. Number of detections and number of occupied sites for each vertebrate taxon during the Autumn 2024 sampling season.

Group	Species	Common name	N detections	N sites
Birds	<i>Acanthizidae</i>	Family of Australian warblers	2	2
	<i>Alectura lathamii</i>	Australian brush-turkey	1	1
	<i>Alisterus scapularis</i>	Australian king-parrot	11	7
	<i>Anatidae</i>	Genus of waterbirds that includes ducks, geese and swans	9	6
	<i>Anthochaera</i>	Genus of honeyeaters and wattlebirds	1	1
	<i>Cacatua</i>	Genus of cockatoos	1	1
	<i>Cacatua galerita</i>	Sulphur-crested cockatoo	13	9
	<i>Chenonetta jubata</i>	Australian wood duck	1	1
	<i>Columba livia</i>	Domestic pigeon	1	1
	<i>Corvus</i>	Genus of crows and ravens	1	1
	<i>Dacelo novaeguineae</i>	Laughing kookaburra	3	3
	<i>Eopsaltria australis</i>	Eastern yellow robin	3	3
	<i>Gallinula tenebrosa</i>	Dusky moorhen	8	5
	<i>Lichenostomus</i>	Genus of honeyeaters	2	2
	<i>Macropygia</i>	Genus of cuckoo-doves	2	2
	<i>Manorina</i>	Genus of honeyeaters	1	1
	<i>Meliphaga lewinii</i>	Lewin's honeyeater	2	2
	<i>Meliphagidae</i>	Family of honeyeaters	4	4
	<i>Menura novaehollandiae</i>	Superb lyrebird	2	1
	<i>Microcarbo melanoleucos</i>	Little pied cormorant	5	4
<i>Ninox</i>	Genus of owls	1	1	
<i>Ninox novaeseelandiae</i>	Morepork	1	1	

Group	Species	Common name	N detections	N sites
	<i>Ninox strenua</i>	Powerful owl	3	2
	<i>Passeriformes</i>	Order of perching birds	7	6
	<i>Phalacrocorax sulcirostris</i>	Little black cormorant	1	1
	<i>Platycercus eximius</i>	Eastern rosella	3	3
	<i>Ptilonorhynchus violaceus</i>	Satin bowerbird	2	1
	<i>Ptilotula penicillata</i>	White-plumed honeyeater	1	1
	<i>Pycnoptilus floccosus</i>	Pilotbird	1	1
	<i>Sericornis frontalis</i>	White-browed scrubwren	2	2
	<i>Trichoglossus haematodus</i>	Rainbow lorikeet	11	8
	<i>Turdus</i>	Genus of true thrushes	1	1
	<i>Zanda</i>	Genus of cockatoos	1	1
	<i>Zosterops lateralis</i>	Silvereye	2	2
	<i>Afurcagobius tamarensis</i>	Tamar goby	2	1
	<i>Aldrichetta forsteri</i>	Yellow-eye mullet	1	1
	<i>Anguilla australis</i>	Shortfin eel	33	23
	<i>Anguilla reinhardtii</i>	Longfin eel	48	28
	<i>Carassius auratus</i>	Goldfish	2	2
	<i>Cyprinus carpio</i>	European carp	9	5
	<i>Galaxias maculatus</i>	Common galaxias	11	7
	<i>Gambusia holbrooki</i>	Eastern gambusia	12	7
	<i>Gobiomorphus australis</i>	Striped gudgeon	20	13
	<i>Gobiomorphus coxii</i>	Cox's gudgeon	37	22
	<i>Hypseleotris</i>	Genus of carp gudgeons	3	2
	<i>Mugil cephalus</i>	Sea mullet	2	1
	<i>Percalates novemaculeata</i>	Australian bass	11	7
	<i>Philypnodon grandiceps</i>	Flatheaded gudgeon	7	4
	<i>Philypnodon macrostomus</i>	Dwarf flathead gudgeon	4	2
	<i>Pseudomugil signifer</i>	Pacific blue-eye	2	1
	<i>Retropinna semoni</i>	Australian smelt	7	4
Fishes & eels				

Group	Species	Common name	N detections	N sites
Frogs	<i>Tandanus tandanus</i>	Eel-tailed catfish, freshwater catfish	3	2
	<i>Trachystoma petardi</i>	Pinkeye mullet	2	1
	<i>Crinia signifera</i>	Common froglet	9	5
	<i>Limnodynastes peronii</i>	Striped marsh frog	3	3
	<i>Litoria phyllochroa</i>	Leaf-green tree frog	7	5
Mammals	<i>Acrobates pygmaeus</i>	Narrow-toed feather-tailed glider	1	1
	<i>Antechinus stuartii</i>	Brown antechinus	2	2
	<i>Bos taurus</i>	Cattle	9	9
	<i>Canis lupus</i>	Dog or dingo	20	17
	<i>Miniopterus</i>	Genus of bats	1	1
	<i>Mus musculus</i>	House mouse	1	1
	<i>Ovis aries</i>	Sheep	2	2
	<i>Perameles nasuta</i>	Long-nosed bandicoot	1	1
	<i>Petaurus breviceps</i>	Sugar glider	7	5
	<i>Pseudocheirus peregrinus</i>	Common ringtail possum	17	14
	<i>Pteropus poliocephalus</i>	Grey-headed flying-fox	10	9
	<i>Rattus</i>	Genus of rats	3	3
	<i>Rattus fuscipes</i>	Bush rat	6	5
	<i>Rattus norvegicus</i>	Brown rat	1	1
	<i>Rattus rattus</i>	Black rat	9	6
	<i>Sus scrofa</i>	Pig	13	10
	<i>Trichosurus vulpecula</i>	Common brush-tailed possum	7	7
	<i>Vulpes vulpes</i>	Red fox	4	4
	<i>Wallabia bicolor</i>	Swamp wallaby	3	3
	Reptiles	<i>Eulamprus quoyii</i>	Eastern water skink	2
<i>Lampropholis delicata</i>		Delicate skink	2	2
<i>Myuchelys latisternum</i>		Saw-shelled turtle	1	1
<i>Scincidae</i>		Genus of skinks	2	2

The figure below shows similar data to those presented in the table above. Rather than focusing on the number of detections, however, this figure shows the percentage of reads assigned to each taxon.



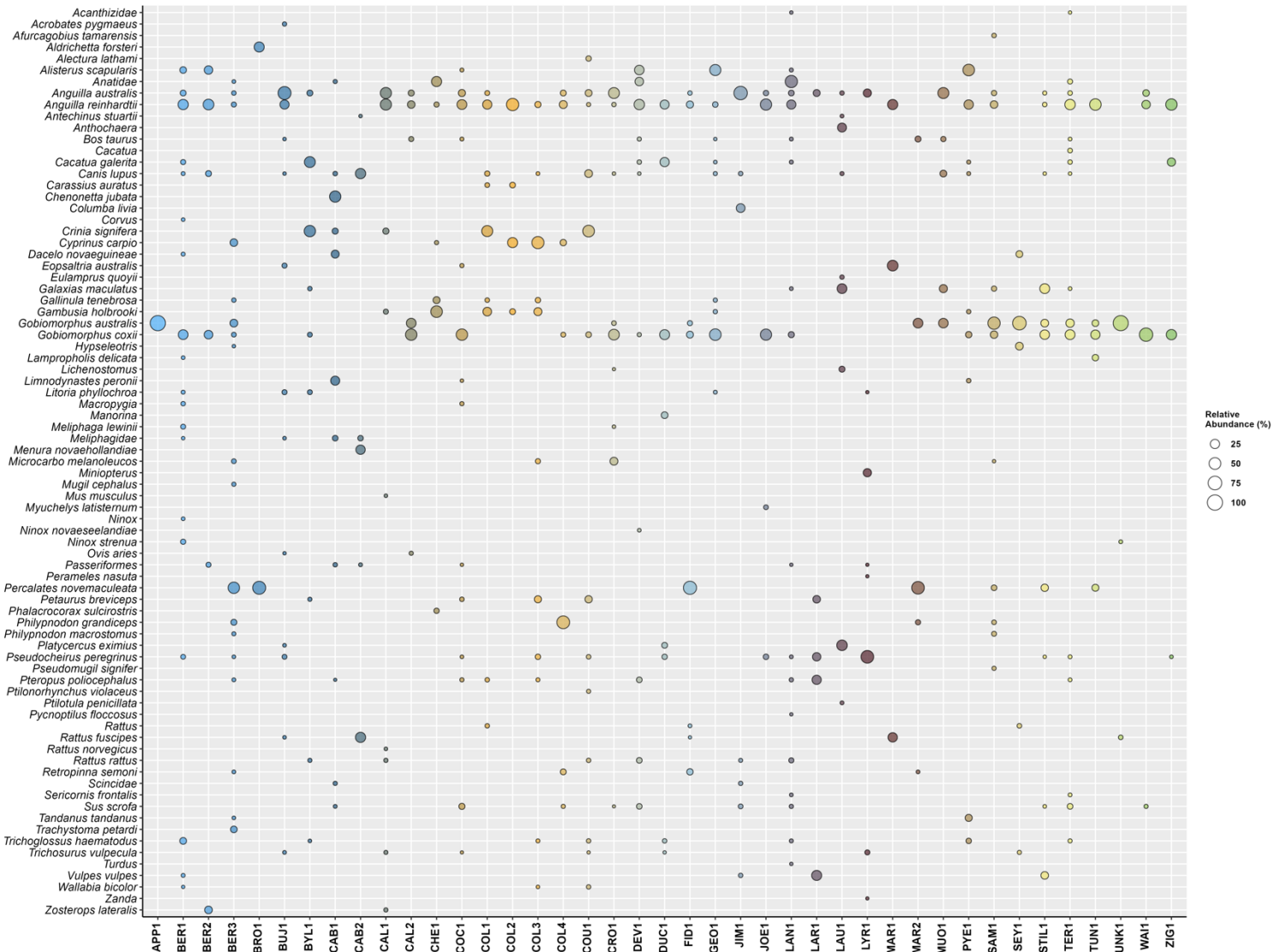


Figure 4. Percentage of reads assigned to each vertebrate taxon at each site in Autumn 2024.

### Decapod Metabarcoding

Overall, five decapod taxa were detected in the Autumn 2024 sampling round. The number of decapod taxa at each site (across all replicate samples) ranged from 0 to 2.

A summary of the frequency of occurrence of each decapod species across the 41 sites is provided in Table 2.

Table 4. Number of detections and number of occupied sites for each decapod taxon in Autumn 2024.

Family	Species	Common name	N detections	N sites
Atyidae	<i>Australatya striolata</i>	Riffle shrimp	3	3
	<i>Cherax destructor</i>	Common yabby	10	7
	<i>Euastacus</i>	Genus of Australian freshwater crayfish	10	9
Parastacidae	<i>Euastacus australasiensis</i>	Sydney spiny crayfish	11	10
	<i>Euastacus spinifer</i>	Giant spiny crayfish	8	5

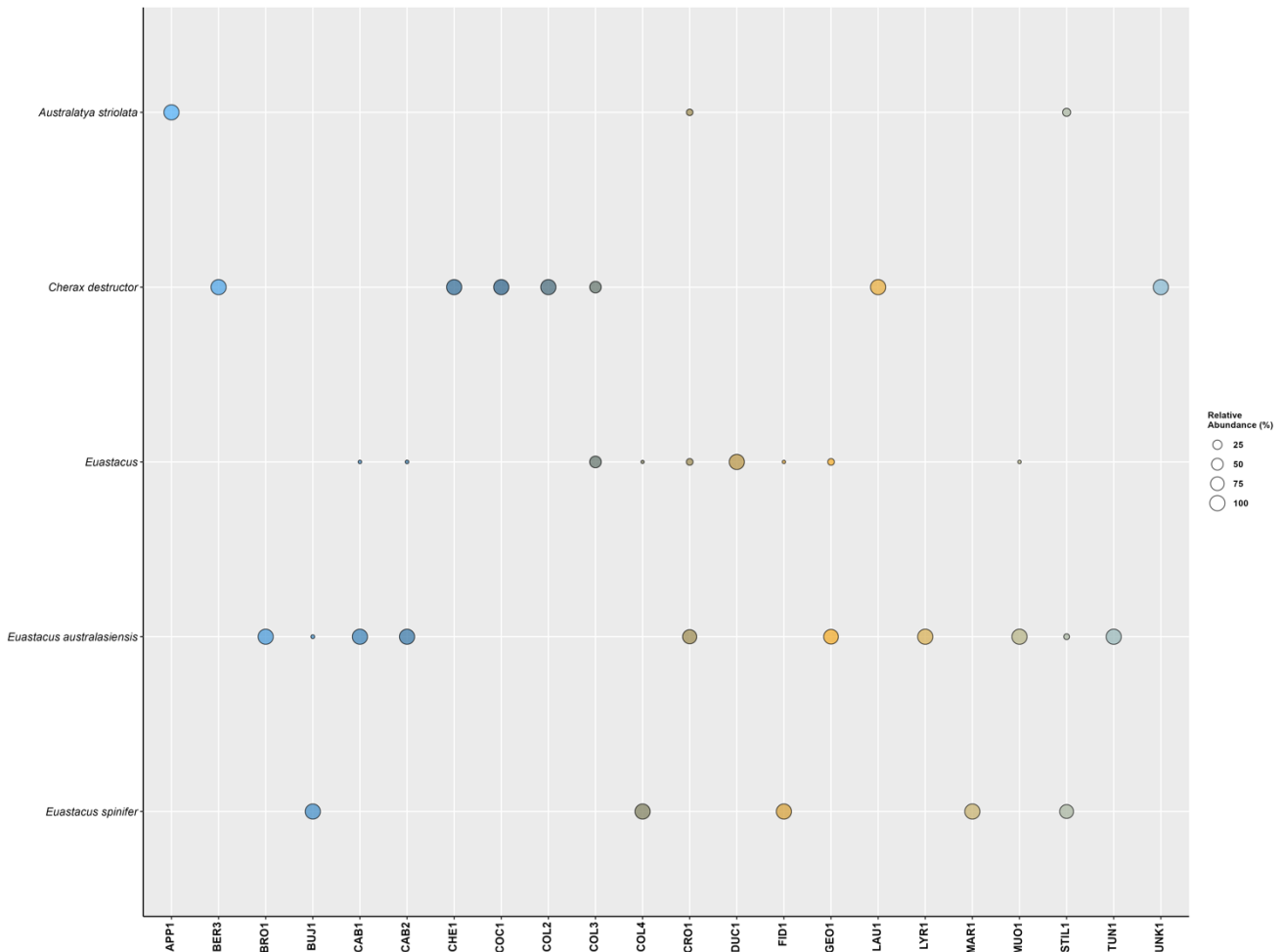


Figure 5. Percentage of reads assigned to each decapod taxon in Autumn 2024.

## Quality Control

- Amplification success was confirmed by gel electrophoresis.
- Numbers of reads in negative controls were below the acceptable threshold.
- All mock community positive controls produced reads of expected species, with no contamination from other species.

### Spring 2023:

- The following controls were used:
  - o 4 extraction controls
  - o 4 mock communities
- The total number of reads was 4,884,394 (vertebrate assay) and 1,138,492 (decapod assay) .
- The median number of reads per sample was 59,458.5 (range = 0 – 92,983) (vertebrate assay) and 2,837 (range = 0 – 114,911) (decapod assay).
- Out of 82 samples analysed, 3 samples were labelled dropouts (fewer than 5,000 non-human reads).

### Autumn 2024:

- The following controls were used:
  - o 12 extraction controls
  - o 4 mock communities
- The total number of reads was 15,432,129 (vertebrate assay) and 9,660,320 (decapod assay).
- The median number of reads per sample was 73,261.5 (range = 0 – 245,534) (vertebrate assay) and 19,514.5 (range = 0 – 687,089) (decapod assay).

## References

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## Disclaimer

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